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Antioxidant Activity and Phenolic and Flavonoid Contents of the Extract and Subfractions of *Euphorbia splendida* Mobayen

Zahra Kefayati^{1,2}, Saeed Mohammadi Motamed¹, Asie Shojaii³, Mitra Noori⁴, Roshanak Ghods⁵

¹Department of Pharmacognosy, Faculty of Pharmacy, Pharmaceutical Sciences Branch, Islamic Azad University, (IAUPS), ²Herbal Medicines Research Center, Pharmaceutical Sciences Branch, Islamic Azad University, Tehran, (HMRC), ³Departments of Traditional Pharmacy and ⁵Traditional Medicine, Research Institute for Islamic and Complementary Medicine and School of Traditional Medicine, Iran University of Medical Sciences, Tehran, ⁴Department of Biology, School of Science, Arak University, Arak, Iran

ABSTRACT

Introduction: The harmful action of the free radicals which cause the oxidative stress can be blocked by antioxidant substances, and different plant extracts showed antioxidant activity. The aim of this study is was evaluation the antioxidant activity of total methanol extract (ME) and subfractions of Euphorbia splendida Mobayen. Materials and Methods: Aerial part of E. splendida was extracted by maceration with methanol and then subfractionated by liquid-liquid fractionation using petroleum ether, chloroform, ethyl acetate, and water. Antioxidant activity was assessed by 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity assay, reduction of ferric ions and ferrous ion chelating potential. Total phenolic contents (TPC) and total flavonoid contents (TFC) were estimated with Folin-Ciocaltue and aluminum chloride methods, respectively. Results: The findings revealed that E. splendida ME and subfractions showed a dose-dependent antioxidant activity. ME showed the highest antioxidant activity based on total reduction capability and ferrous ions chelating assay tests. Aqueous fraction and then ethyl acetate fraction showed the best $\mathrm{IC}_{_{50}}$ in DPPH radical scavenging test in comparison to butylated hydroxytoluene. ME showed the highest value of TPC and TFC (270.74 ± 0.005 mg/g and 208.23 ± 0.007 mg/g, respectively). Conclusion: This study showed that the extract and subfractions of E. splendida have antioxidant activity. The antioxidant activity of the extract and fractions might be attributed to the presence of phenolic compounds. More studies are needed to determine the active antioxidant compounds of this plant.

Key words: 1,1-Diphenyl-1-picrylhydrazyl, antioxidant, *Euphorbia splendida,* flavonoid, fractions, free radicals

SUMMARY

• Total extract and subfractions of Euphorbia splendida showed antioxidant activity.

Abbreviations Used: TPC: Total phenolic content, TFC: Total flavonoid content, DPPH: 2, 2'- diphenyl-1-picrylhydrazyl, BHT: Butylated hydroxytoluene, EDTA: Ethylene Diamine Tetra Acetic acid, ME: Total methanol extract, EAF: Ethyl acetate fraction , AQF: Aqueous fraction, PEF: Pertolium ether fraction, CHF: Chloroformic fraction

Ferric reducing Extract DPPH(IC50) Chelating & activity (%)3 $(\mu g/ml)^2$ power (nm)4 fractions 1 0.544±0.004 ME 145.47±0.007 79±3.1 PEF 482.99±0.009 0.491±0.008 61.2±3.96 0.519±0.012 CHF 287.14±0.10 68 5+4 65 EAF 134.28±0.005 68±6.21 0.518±0.01 AQF 129.02±0.006 75±3.85 0.514±0.01 BHT 78.23±0.1 EDTA 99.07 ± 0.49

Euphorbia splendida extract and fractions

¹ME= Total methanolic extract, PEF=pertolium ether fraction, EAF= Ethyl acetate fraction, AQF= Aquoes fraction, BHT= Butylated hydroxy toluene, EDTA= **Ethylene diamine tetra acetic acid**.

²The concentration (µg/ml) of the plant extracts for inhibition of 50% DPPH free radicals (IC50).

³ The percentage of iron chelating activity of the plant extracts in the concentration of 1000 µg/ml.

⁴ Increased control absorbance by the plant extracts (600 μg/m1/(increased control absorbance by BHT 1000)

µg/ml) × 100

Correspondence:

Dr. Asie Shojaii, Department of Traditional Pharmacy, Research Institute for Islamic and Complementary Medicine and School of Traditional Medicine, Iran University of Medical Sciences, Vahdat Eslami Street, Behesht Street, Tehran, Iran. E-mail: shojaii.a@iums.ac.ir **DOI:** 10.4103/pr.pr_12_17





INTRODUCTION

It has been shown that oxidative stress is one of the major causative factors in the induction of many chronic and degenerative diseases including atherosclerosis, ischemic heart disease, aging, diabetes mellitus, cancer, neurodegenerative diseases, and others.^[1] The harmful action of the free radicals which cause the oxidative stress can be blocked by antioxidant substances, which scavenge the free radicals and detoxify the organism.^[2] Several plant extracts and different classes of phytochemicals have been shown to have antioxidant activity.^[3,4] This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

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Genus *Euphorbia* (Euphorbiaceae) comprising about 2000 species and spreads in Pakistan, India, and Iran and over 82 species of *Euphorbia* have been found in Iran.^[5] Previous studies on *Euphorbia* species or their active components showed different biological effects such as cytotoxic, antitumor, antioxidant, antibacterial,^[6] anti-inflammatory, and antinociceptive activities.^[7-9] Up to now, antioxidant activity of some species of *Euphorbia* such as *Euphorbia helioscopia* and *Euphorbia hirta* has been reported.^[10,11]

Euphorbia splendida Mobayen is a plant distributed in the West of Iran with 30–50 cm height.^[12] our previous studies on *E. splendida* showed the presence of diterpenoid, triterpenoid, and flavonoid in this plant.^[13,14] According to our investigation, antioxidant activity of this plant has not been studied so far. Therefore, in this study, antioxidant activity of the methanol extract (ME) and subfractions of *E. splendida* has been evaluated with different methods.

MATERIALS AND METHODS

Plant material

Fresh aerial parts of *E. splendida* were collected in May 2014 from Arak, Markazi province, Iran. The specimen was identified by Dr. M. Noori (the Biology Department, Faculty of Science, Arak University) and was deposited under voucher number CMK10 in the herbarium of the biology department.

Extraction and isolation

The aerial parts of plant (600 g) were dried, ground, and extracted with methanol by maceration. The extraction was repeated for three times (3 days for each time) in 25°C. Different fractions of the extract were obtained by liquid–liquid fractionation using water, petroleum ether (PE), chloroform, and ethyl acetate (1500 cc of each solvent). The extract and fractions were concentrated by rotary evaporator (IKA, Model, RV 10 D) and finally dried and stored in a clean, dark container and cool place. The antioxidant activity of the extract and fractions was determined using 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity, total reduction capability, and ferrous ions chelating assays.

Chemicals

DPPH, butylated hydroxytoluene (BHT), and gallic acid were purchased from Sigma-Aldrich USA. Folin–Ciocalteu was obtained from Merck (Darmstadt, Germany). Potassium ferricyanide, potassium acetate, phosphate buffer, ferrous ammonium sulfate, ascorbic acid, aluminum chloride (AlCl₃), thrichloroacetic acid (TCA), ammonium molybdate, tannic acid, quercetin, acetyl acetone, and ferric chloride (FeCl₃) were purchased from Sigma Chemical Co. (St. Louis, MO, USA). Rutin, gallic acid, TCA, potassium ferricyanide, ferrozine, and BHT were purchased from Sigma, and iron (II) chloride was purchased from Aldrich. Methanol, chloroform, ethyl acetate, and PE were purchased from Merk (Darmstadt, Germany).

Total flavonoid content

The total flavonoid content (TFC) of *E. splendida* ME and subfractions was determined using $AlCl_3$ reagent.^[15] Briefly, 2.5 mL of each sample (and/or rutin as the standard), previously dissolved in 90% ethanol, was mixed with 2.5 mL of $AlCl_3 2\%$ solution in 90% ethanol. After 40 min, the absorbance of the produced yellow solution was measured at 425 nm. The TFC of the samples was calculated on the basis of a linear calibration curve obtained using rutin.

Total phenolic content

The content of total phenolic compounds in plant extract and fractions was determined by Folin–Ciocalteu method.^[16] One milliliter of plant extract (concentration of 1 mg/mL) and fractions was dissolved

in methanol and mixed with 5 mL Folin–Ciocalteu reagent and 4 mL (7.5 g/100 mL) sodium carbonate. After 1 h at room temperature, the absorption of clear solutions was read at 765 nm. For the preparation of calibration curve, different concentrations of gallic acid solution were mixed with the same reagents as described above. The amount of total phenolics was expressed as gallic acid equivalent (GAE) in milligrams per gram dry plant extract.

1,1-Diphenyl-2-picrylhydrazyl free radical scavenging assay

The free radical scavenging activity of the ME and subfractions was evaluated using DPPH methods.^[17] Briefly, 1 mL of the sample solution with different concentrations (ranging from 50 to 1000 μ g/mL) was mixed with 3 mL of DPPH methanol solution. The reaction mixtures were incubated at room temperature and allowed to react for 30 min in the dark. After 30 min, the absorbance values were measured at 517 nm and converted into a percentage of antioxidant activity. BHT was used as a positive standard control. The percentage inhibition of DPPH (%) was calculated as follows:

Inhibition of DPPH % =
$$\frac{ODc - (ODs - ODs)}{ODc} \times 100$$

ODc = Control solution absorbance.

 $OD_{b} = Blank$ solution absorbance.

 $OD_s = Sample solution absorbance.$

The concentration of sample required to scavenge 50% of the DPPH free radicals (IC_{50}) was determined from the curve of percentage inhibitions plotted against the respective concentration.

Ferric reducing antioxidant power assay

The total antioxidant potential of the ME and subfractions of *E. splendida* was determined according to method of Oyaizu.^[18] Aliquot (0.25 mL) of samples solution at different concentrations (ranging from 25 to 600 µg/mL) was mixed with 2.5 mL of phosphate buffer (pH 6.6) and 2.5 mL of 1% (w/v) solution of potassium ferricyanide. Then, all the mixtures were incubated in a water bath at 50°C for 20 min. Then, 2.5 mL of 10% (w/v) TCA solution was added and the mixture was then centrifuged at 3000 rpm for 10 min. A volume of 2.5 mL of the supernatant was combined with 2.5 mL of distilled water and 0.5 mL of a 0.1% (w/v) solution of FeCl₃. The absorbance was measured at 700 nm with a spectrophotometer uv-vis (UNICO Model, UV/VIS 2100). BHT was used as positive control. All the tests were done in triplicate and results were reported as mean \pm standard deviation.

Metal chelating activity

The chelating of ferrous ions by the ME and subfractions of *E. splendida* was estimated by the method of Dinis *et al.*, 1994.^[19]

Table 1: Total flavonoid content and total phenolic content of Euphorbia
splendida extract and fractions (data are mean±standard deviation)

Extract and fractions	TPC (mg/g)	TFC (mg/g)
ME	270.74±0.005	208.23±0.007
PEF	101.43±0.007	93.84±0.006
CHF	235.63±0.006	172.75±0.005
EAF	208.54±0.010	65.80±0.006
AQF	173.88±0.005	85.28±0.010

ME: Total methanolic extract; PEF: Petroleum ether fraction; CHF: Chloroformic fraction; EAF: Ethyl acetate fraction; AQF: Aqueous fraction; TPC: Total phenolic content as mg/g plant sample in gallic acid equivalent; TFC: Total flavonoid content as mg/g plant sample in rutin equivalent Briefly, 1 mL of each test sample (1 mg/mL) was taken and added to 0.5 mL of 2 mM FeCl_2 . The reaction was initiated by the addition of 0.2 mL of 5 mM ferrozine into the mixture, which was then left at room temperature for 10 min and then the absorbance of the mixture was read at 562 nm.

RESULTS

Total flavonoid content and total phenolic content of the extract and subfractions of *Euphorbia splendida*

The total phenolic content (TPC) of the ME and subfractions calculated from regression equation of calibration curve ([y = 5.0121x, $R^2 = 0.99$]) and expressed in GAEs, varied between 101.43 and 270.74 mg GAE/g plant sample [Table 1]. The content of total flavonoids in plant samples (mg/g) calculated from regression equation of calibration curve (y = 5.3267x, $R^2 = 0.99$) was expressed in rutin equivalents (REs) varied between 65.80 and 208.23 mg RE/g plant sample [Table 1]. Total methanolic extract showed the highest TFC and TPC values.

DPPH free radical scavenging assay

ME of the aerial parts of *E. splendida* and aqueous, ethyl acetate, chloroform, and PE subfractions were analyzed for DPPH radical

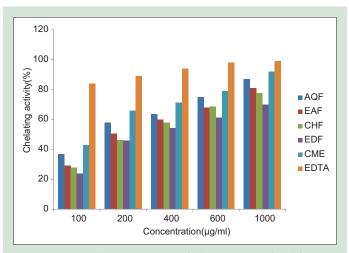


Figure 1: Iron chelating activity of the extract and subfractions of *Euphorbia splendida*. EDTA: Ethylenediaminetetraacetic acid; CME: Methanol extract; EDF: Petroleum ether fraction; CHF: Chloroform fraction; EAF: Ethyl acetate fraction; AQF: Aqueous fraction

scavenging activity to obtain their concentrations to scavenge 50% DPPH (IC₅₀) as shown in Table 2. The aqueous and then ethyl acetate subfractions of *E. splendida* were shown the best results in inhibition of DPPH radical in comparison to standard (BHT). PE subfraction was shown the least activity with IC₅₀ = 482.99 ± 0.01 µg/mL.

Metal chelating activity

Metal chelating activity (%) of the plant ME, subfractions, and ethylenediaminetetraacetic acid (EDTA) in concentrations of 100, 200, 400, 600, and 1000 µg/mL are shown in Figure 1. The ME and subfractions of *E. splendida* showed a dose-dependent antioxidant activity in this method comparing to EDTA. In concentration of 1000 µg/mL, ME showed the strongest activity ($79\pm3.07\%$) and petroleum ether fraction (PEF) showed the lowest activity ($61.2\pm3.96\%$) [Table 2].

Ferric reducing antioxidant power assay

In ferric reducing power assay, all the samples increased the absorbance of the control solution (0.210 ± 0.098) in 700 nm. As shown in Figure 2, all the concentrations had lower absorbance and consequently lower reduction capability toward BHT.

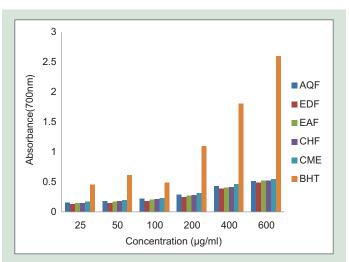


Figure 2: Ferric reducing power activity of the extract and subfractions of *Euphorbia splendida*. Standard; butylated hydroxytoluene. All the samples showed activity lower than butylated hydroxytoluene. Control absorbance: 0.210 ± 0.098 . CME: Methanol extract; EDF: Petroleum ether fraction; CHF: Chloroform fraction; EAF: Ethyl acetate fraction; AQF: Aqueous fraction

Table 2: Radical scavenging activity in the 2, 2'- diphenyl-1-picrylhydrazyl assay, metal chelating activity, and ferric reducing power activity of methanol extract and subfractions of *Euphorbia splendida* (mean±standard deviation)

Extract and fractions	DPPH (IC _{₅0}) (µg/ml)ª	Chelating activity (%) ^b	Ferric reducing power (nm) ^c
ME	145.47±0.007	79±3.1	0.544 ± 0.004
PEF	482.99±0.009	61.2±3.96	0.491 ± 0.008
CHF	287.14±0.10	68.5±4.65	0.519±0.012
EAF	134.28±0.005	68±6.21	0.518 ± 0.01
AQF	129.02±0.006	75±3.85	0.514 ± 0.01
BHT	78.23±0.1	-	2.6±0.1
EDTA	-	99.07±0.49	-

^eThe concentration (μ g/ml) of the plant extracts for inhibition of 50% DPPH free radicals (IC_{50}); ^bThe percentage of iron chelating activity of the plant extracts in the concentration of 1000 μ g/ml; ^cThe absorbance of the plant samples (nm) in concentration of 600 μ g/ml. ME: Total methanolic extract; PEF: Petroleum ether fraction; CHF: Chloroformic fraction; EAF: Ethyl acetate fraction; AQF: Aqueous fraction; BHT: Butylated hydroxytoluene; EDTA: Ethylenediaminetetraacetic acid; DPPH: 2, 2'- diphenyl-1-picrylhydrazyl

DISCUSSION

Several plant extracts and different classes of phytochemicals have been shown to have antioxidant activity. Different species in genus *Euphorbia* showed antioxidant and free radical scavenging activity.^[10,11] *E. splendida* is a plant which contains different chemical compounds,^[13,14] and until now, it is not studied for antioxidant activity. Therefore, in the present study, antioxidant activity of the ME and subfractions of this plant was studied using DPPH free radical scavenging assay, metal chelating activity, and ferric reducing antioxidant power assay.

In DPPH method which is a good method to evaluate radical scavenging activity of the plants, the potency of *E. splendida* ME and subfractions was as below:

Aqueous fraction (AQF) > ethyl acetate fraction (EAF) > Total methanol extract (ME) > chloroformic fraction (CHF) > petroleum ether fraction (PEF).

The free radical scavenging activity was expressed as the effective concentration required for 50% of DPPH radical (DPPH) reduction (IC₅₀) obtained from a plot of graph of scavenging activity against the concentration of the extract and its fractions. The AQF showed the highest activity (IC₅₀ = 129.02 ± 0.01 µg/mL) compared to other extract and fractions which was lower than IC₅₀ of BHT (78.23 ± 0.1 µg/mL).

For the other tests, the order of potency of ME and subfractions was as below:

- Metal chelating activity: ME > AQF > CHF > EAF > PEF
- Total reduction capability: ME > CHF > EAF > AQF > PEF
- Phenolic contents: ME > CHF > EAF > AQF > PEF
- Flavonoid contents: ME > CHF > PEF > AQF > EAF.

The obtained results for total reduction capability are in agreement with the TPCs determined for extract and subfractions. There are acceptable correlation coefficients (R) between phenolic content and the data of DPPH scavenging activity, metal chelating activity, and total reduction capability of the plant extract and subfractions (R = -0.68, 0.73, and 0.95,respectively); such correlation with flavonoid content was R = -0.06, 0.51, and 0.69, respectively, which was not as significant as phenolic contents. Different species of Euphorbia showed antioxidant activity in different antioxidant assays.^[10,20] Comparing to E. splendida, some of Euphorbia species such as *E. hirta* showed lower IC₅₀ value in DPPH scavenging assay which was comparable to standards such as BHT.^[21] Hence, although E. splendida showed significant antioxidant activity in comparison to standards, especially in DPPH and metal chelating activity, this antioxidant activity may be lower than antioxidant activity of some Euphorbia species such as E. hirta. The extract and subfractions of this plant are found to have different levels of antioxidant activity and phenolic and flavonoid contents. There are some flavonoids such as quercetin and rutin which have been reported from E. splendida.^[14] These flavonoids showed significant antioxidant activity in different assays.^[22-24] Plant polyphenols act as reducing agents and antioxidants by the hydrogen-donating property of their hydroxyl groups,^[25] so these polyphenols may be responsible for the observed antioxidant activity. More studies are recommended to determine the active antioxidant compounds of this plant.

CONCLUSION

This study showed that the extract and subfractions of the aerial parts of E. splendida have antioxidant activity. The antioxidant activity of the extract and fractions might be attributed to the presence of phenolic compounds. More studies are needed to determine the active antioxidant compounds of this plant.

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Nil.

Conflicts of interest

There are no conflicts of interest.

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