

Effect on non-enzymatic antioxidants

GSH, total thiols and vitamin C levels of nitrobenzene-induced and ethanol extract treated groups (group II and group III) are shown in Table 4. The levels of GSH, total thiols and vitamin C were significantly ($P < 0.01$) decreased in the nitrobenzene-induced group (group II) as compared to the normal group (group I). On treatment with the ethanol extract of *E. hirta* (group III), GSH, total thiols and vitamin C levels were found to be enhanced significantly ($P < 0.01$). There was no significant difference between the ethanol extract alone treated group (group IV) and normal group (group I).

Histopathology

Histopathologic studies also provided a supportive evidence for biochemical analysis. In case of control (group I), the kidney section showed normal glomeruli, Bowman's tubules and blood vessels. The kidney section of the animals induced with nitrobenzene (group II) showed glomerular degeneration with loss of surrounding Bowman's capsule and tubular necrosis. Treatment with the ethanol extract of *E. hirta* (group III) showed a glomerulus with an intact Bowman's capsule, and all the renal cells which were damaged due to nitrobenzene appeared to be regenerated and almost showed normal histology. The kidney section of the animals that received the ethanol extract of *E. hirta* alone (group IV) showed normal morphology [Figure 1].

DISCUSSION

Blood urea nitrogen (BUN), serum urea, uric acid and

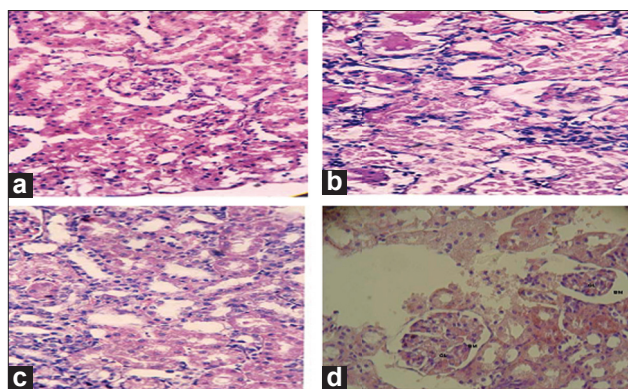


Figure 1: Histopathology of kidney. (a) Kidney section of the normal rat (group I) showing normal tubular brush borders and intact glomerulus and surrounding Bowman's capsule; (b) kidney section of the rat induced with nitrobenzene (group II) showing a glomerulus with loss of surrounding Bowman's capsule and tubular necrosis with swelling and vacuolation; (c) kidney section of the rat induced with nitrobenzene and treated with ethanol extract of *Euphorbia hirta* (group III) showing a glomerulus with an intact Bowman's capsule and mildly swollen tubules and almost showing normal morphology; (d) kidney section of rat treated with extract alone showing normal glomerulus and Bowman's capsule

creatinine are considered as the indicators of kidney damage, and their abnormal levels indicate marked renal injury. In our previous basic study, nitrobenzene induction produced an elevation in the concentrations of BUN, serum urea, and creatinine and significant reduction in the level of uric acid. Treatment with *E. hirta* significantly restored the levels to normalcy.^[32] In the present study, nitrobenzene increased the level of renal tissue MDA production, which suggested involvement of free radicals in nitrobenzene-induced nephrotoxicity. Nitrobenzene produces a number of free radicals during its reactive metabolism in the gut as well as at the cellular level and generates superoxide anion as a by-product during oxidative metabolism. The reactive species generated during nitrobenzene metabolism are considered candidates for carcinogenicity. Furthermore, several lines of evidence suggest that nitrobenzene exerts its carcinogenicity through a non-DNA reactive (epigenetic) fashion, such as a strong temporal relationship between non-, pre- and neoplastic lesions leading to carcinogenesis.^[33]

The elevated levels of LPO in nitrobenzene-induced animals might be due to the renal damage caused by nitrobenzene-induced free radical generation. LPO, an index of oxidative stress, leads to deterioration of biological system^[34] and is ascribed to a free radical mediated chain reaction that damages cell membrane and implicates renal oxidative stress.^[35] Oxidative stress is believed to be a primary factor in various diseases.^[36] Oxidative stress could also be involved in other inflammatory glomerular lesions caused by a series of mediators including cytokines and chemokines, which leads to leukocyte activation, production of reactive oxygen species (ROS) and increased glomerular damage.^[37] Different studies have shown that free radicals are responsible for oxidative damage to different molecules (lipids, proteins, nucleic acid) and thus are involved in the initiation phase of some degenerative illnesses.^[38] The elevated level of lipid peroxides in nitrobenzene-induced group would be a transient phenomenon of cellular damage.^[39] In animals treated with ethanol extract of *E. hirta*, the rise in lipid peroxides was prevented significantly. The decrease in lipid peroxides may be due to the antioxidant effect of the extract.

The generation of ROS in kidney has been proposed as a mechanism by which many chemicals can induce nephrotoxicity.^[40] The ROS generation in tissues is efficiently scavenged by the enzymatic and non-enzymatic antioxidants. The decrease in the activities of antioxidant enzymes is in close relationship with the induction of LPO.^[41] Thus, the mechanism of nephrotoxicity is related to the depletion of the antioxidant defense system.^[1] The reduced activity of SOD, CAT, GPx and GST could be

due to enhanced LPO or inactivation of the antioxidative enzymes.^[42] The present work confirms that the observed decrease in the antioxidant activity was presumably associated with the increased LPO and oxidative stress caused by nitrobenzene. The ethanol extract administration restored the depleted renal antioxidants. This recovery to near normalcy reveals that oxidative stress induced by nitrobenzene has been nullified due to the antioxidant potential effect of *E. hirta*.

The antioxidant activity may be due to the inhibition of the formation of radicals or scavenging of the formed radical.^[43] Supplementation of plant material improves all these parameters. Similar results have been reported by a number of investigators.^[44,45] The biochemical findings were also supported by the histopathologic studies which showed a glomerulus with loss of surrounding Bowman's capsule and diffuse tubular necrosis in the renal section of rats induced with nitrobenzene. These histopathologic changes almost disappeared in the renal tissue of rats treated with the ethanol extract of *E. hirta*. Numerous studies have also shown that medicinal plants protect the kidney against nephrotoxicity induced by various chemicals.^[46,47] Thus, the ethanol extract of *E. hirta* at a dosage of 400 mg/kg body weight showed a significant change against nitrobenzene-induced nephrotoxicity. Preliminary phytochemical screening of the ethanol extract of *E. hirta* showed a higher concentration of flavonoid. Literature has shown medicinal plants with nephroprotective properties to mediate their protection via antioxidant and/or free radical scavenging activities due to the high concentration of flavonoids and alkaloids that they contain.^[48,49] Thus, in the present study, nephroprotection offered by the ethanol extract of *E. hirta* could be due to the presence of higher concentration of flavonoids present in it.

CONCLUSION

Thus, the results of this investigation suggest that the ethanol extract of *E. hirta* can prevent the toxic effects of nitrobenzene and can be used in the treatment of nephrotoxicity. The protective action of the ethanol extract of *E. hirta* was probably due its antioxidant nature which protects the kidney against nephrotoxicity induced by nitrobenzene.

ACKNOWLEDGMENT

We, the authors, are thankful to Chancellor, Advisor, Vice Chancellor and Registrar of Karpagam University for providing facilities and encouragement.

REFERENCES

- Joy J, Nair CK. Amelioration of cisplatin induced nephrotoxicity in Swiss albino mice by *Rubia cordifolia* extract. J Can Res Ther 2008;4:111-5.
- Wojcikowski K, Stevenson L, Leach D, Wohlmuth H, Gobe G. Antioxidant capacity of 55 medicinal herbs traditionally used to treat the urinary system: A comparison using a sequential three solvent extraction process. J Altern Complement Med 2007;13:103-9.
- James W, Joint J, Siegel C, Singh D, Velazquez S. Nitrobenzene carcinogenicity, National Center for Environmental Assessment. 1998; EPA/600/R-95/100.
- Cattley RC, Everitt JI, Gross EA, Moss OR, Hamm TE Jr, Popp JA. Carcinogenicity and toxicity of inhaled nitrobenzene in B6C3F1 mice and F344 and CD rats. Fundam Appl Toxicol 1994;22:328-40.
- Howard PC, Beland FA, Cerniglia CE. Reduction of the carcinogen 1- nitropyrene to 1 - aminopyrene by the rat intestinal bacteria. Carcinogenesis 1983;4:985-90.
- Montilla P, Barcos M, Munoz MC, Bujalance I, Munoz-Castaneda JR, Tunez I. Red wine prevents brain oxidative stress and nephropathy in streptozotocin-induced diabetic rats. J Biochem Mol Biol 2005;38:539-44.
- Mansour HH, Hafez HF, Fahmy NM. Silymarin modulates cisplatin-induced oxidative stress and hepatotoxicity in rats. J Biochem Mol Biol 2006;39:656-61.
- El-Beshbishy HA. Hepatoprotective effect of green tea (*Camellia sinensis*) extract against tamoxifen-induced liver injury in rats. J Biochem Mol Biol 2005;38:563-70.
- Umamaheswari M, Chatterjee TK. Effect of the fractions of *Coccinia grandis* on Ethanol-Induced cerebral oxidative stress in rats. Phcog Res 2009;1:25-34.
- Samarghandian S, Afshari JT, Davoodi S. Honey induces apoptosis in renal cell carcinoma. Phcog Mag 2011;7:46-52.
- Shirwaikar A, Verma R, Lobo R. Phytotherapy - Safety aspects. Natural Product Radiance, 2009;8:55-63.
- Shanmugam KR, Ramakrishna CH, Mallikarjuna K, Reddy KS. Protective effect of ginger against alcohol-induced renal damage and antioxidant enzymes in male albino rats. Indian J Exp Biol 2010;48:143-9.
- Chidrawar VR, Ushir YV, Sudarshan S, Patel KN, Patel NJ, Vadalia KR. Possible Role of Natural Nephroprotective; *Hemidesmus indicus* in Congestive Heart Failure. Phcog Res 2009;1:367-74.
- Abubakar EM. Antibacterial activity of crude extracts of *Euphorbia hirta* against some bacteria associated with enteric infections. J Med Plants Res 2009;3:498-505.
- Ogbulie JN, Ogueke CC, Okoli IC, Anyanwu BN. Antibacterial activities and toxicological potentials of crude ethanol extracts of *Euphorbia hirta*. Afr J Biotechnol 2007;6:1544-8.
- Sharma NK, Dey S, Prasad R. *In vitro* antioxidant potential evaluation of *Euphorbia hirta*. Pharmacol Online 2007;1:91-8.
- Kumar S, Malhotra R, Kumar D. *Euphorbia hirta*: Its chemistry, traditional and medicinal uses, and pharmacological activities. Phcog Rev 2010;4:58-61.
- Liu Y, Murakami N, Ji H, Abreu Pedro, Zhang S. Antimalarial flavonol glycosides from *Euphorbia hirta*. Pharm Biol 2007;45:278-81.
- Adedapo AA, Abatan MO, Idowu SO, Olorunsogo OO. Effects of chromatography fractions of *Euphorbia hirta* on the rat serum biochemistry. Afr J Biomed Res 2005;8:185-9.
- Kandalkar A, Patel A, Darade S, Baviskar D. Free radical scavenging activity of *Euphorbia hirta* Linn. leaves and

- isolation of active flavonoid myricitrin. *Asian J Pharm Clin Res* 2010;3:234-37.
21. Khan MR, Rizvi W, Khan GN, Khan RA, Shaheen S. Carbon tetrachloride-induced nephrotoxicity in rats: Protective role of *Digera muricata*. *J Ethnopharmacol* 2009;122:91-9.
 22. Trease GE, Evans, WC. A textbook of Pharmacognosy. London: Bacilliere Tinnall Ltd; 1989.
 23. Lowry OH, Roseberg NJ, Farr AL, Randell RJ. Protein measurement with Folin phenol reagent. *J Biol Chem* 1951;193:265-75.
 24. Hogberg J, Larson RE, Kristoferson A, Orrhenices S. NADPH-dependent reductase solubilised from microsomes by peroxidation and its activity. *Biochem Biophys Res Commun* 1974;56:836-42.
 25. Misra HP, Fridovich I. The role of superoxide anion in the antioxidant epinephrine and a single assay of superoxide dismutase. *J Biol Chem* 1972;247:3170-5.
 26. Lueck H. *Methods of enzymatic analysis*. London: Academic Press; 1965.
 27. Rotruck JT, Pope AL, Ganther HS. Selenium: Biochemical role as a component of glutathione peroxidase purification and assay. *Science* 1973;179:588-90.
 28. Habig WH, Pabst MJ, Jakoby WB. Glutathione-S-transferases. The first enzymatic step in mercapturic acid formation. *J Biol Chem* 1974;249:7130-9.
 29. Moran MS, Diferre JW, Manneruik B. Levels of glutathione reductase glutathione-s-transferase in rat lung and liver. *Biochim Biophys Acta* 1979;582:67-78.
 30. Sedlack J, Lindsay RH. Estimation of total protein bound non protein bound sulphhydryl group in the tissue with Ellmann's reagent. *Anal Biochem* 1968;25:192-205.
 31. Omaye ST, Turabull JD, Sauberlich HE. Selected method for the determination of ascorbic acid in animal cells tissues and fluids. *Methods Enzymol* 1979;62:3-11.
 32. Suganya S, Ragavendran P, Rajalakshymenon B, Rathi MA, Thirumoorthi L, Meenakshi P, *et al.* Potential effect of *Euphorbia hirta* against nitrobenzene-induced nephrotoxicity. *Pharmacol Online* 2010;2:963-70.
 33. Hsu CH, Stedeford T, Okochi-Takada E, Ushijima T, Noguchi H, Muro-Cacho C, *et al.* Framework analysis for the carcinogenic mode of action of nitrobenzene. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev* 2007;25:155-84.
 34. Ray S, De K, Sengupta C, Roy K. QSAR study of lipid peroxidation-inhibition potential of some phenolic antioxidants. *Indian J Biochem Biophys* 2008;45:198-205.
 35. Noori S, Mahboob T. Antioxidant effect of carnosine pretreatment on cisplatin induced renal oxidative stress in rats. *Indian J Clin Biochem* 2010;25:86-91.
 36. Bhujbal SS, Kewatkar SM, More LS, Patil MJ. Antioxidant Effects of Roots of *Clerodendrum serratum* Linn. *Phcog Res* 2009;1:294-8.
 37. Ragheb A, Attia A, Eldin WS, Elbarbry F, Gazarin S, Shoker A. Protective effect of thymoquinone, an anti-oxidant, an anti-inflammatory agent against renal injury: A review. *Saudi J Kidney Dis Transpl* 2009;20:741-52.
 38. Stef DS, Gergen I, Trasea TI, Harmanescu M, Lavinia S, Ramona MB, *et al.* Total antioxidant and radical scavenging capacities for different medicinal herbs. *Rom Biotechnol Lett* 2009;14:4704-9.
 39. Bhartiya US, Raut YS, Joseph L. Protective effect of *Ocimum sanctum* L after high dose of ¹³¹Iodine exposure in mice as *in vivo* study. *Indian J Exp Biol* 2006;44:647-52.
 40. Somani SM, Husain K, Whitworth C, Trammell GL, Malafa M, Rybak LP. Dose-dependent protection by lipoic acid against cisplatin-induced nephrotoxicity in rats: Antioxidant defense system. *Pharmacol Toxicol* 2000;86:234-41.
 41. Chandra Jagetia G, Rajanikant GK, Rao SK, Shrinath Baliga M. Alteration in glutathione, glutathione peroxidase, superoxide dismutase, and lipid peroxidation by ascorbic acid in the skin of mice exposed to fractionated gamma radiation. *Clin Chim Acta* 2003;332:111-21.
 42. Rahman I, MacNee W. Lung glutathione and oxidative stress: Implications in cigarette smoking-induced airway disease. *Am J Physiol* 1999;277:1067-88.
 43. Gutierrez RMP, Navarro YTG. Antioxidant and hepatoprotective effects of the methanol extract of the leaves of *Satureja macrostema*. *Phcog Mag* 2010;6:125-31.
 44. Devi DG, Lija Y, Cibin TR, Biju PG, Devi VG, Abraham A. Evaluation of the protective effects of *Emilia sonchifolia* Linn. (DC.) on perchlorate induced oxidative damage. *J Biol Sci* 2006;6:887-92.
 45. Nwanjo HU, Oze G, Okafor MC, Nwosu D, Nwankpa P. Oxidative stress and non-enzymatic antioxidant status in hypertensive patients in Nigeria. *Afr J Biotechnol* 2001;6:1681-4.
 46. Priya R, Vasudha KC. Antioxidant vitamins in chronic renal failure. *Biomed Res* 2009;20:67-70.
 47. Sinha M, Manna P, Sil PC. Amelioration of galactosamine-induced nephrotoxicity by a protein isolated from the leaves of the herb *Cajanus indicus* L. *BMC Complement Altern Med* 2007;7:11.
 48. Ajamia M, Eghtesadia S, Pazoki-Toroudib H, Habibeyb CR, Ebrahimid SA. Effect of *Crocus sativus* on gentamicin induced nephrotoxicity. *Biol Res* 2010;43:83-90.
 49. Ghosh A, Sil PC. Anti-oxidative effect of a protein from *Cajanus indicus* L. against acetaminophen-induced hepato-nephro toxicity. *J Biochem Mol Biol* 2007;40:1039-49.

Cite this article as: Suganya S, Sophia D, Raj CA, Rathi MA, Thirumoorthi L, Meenakshi P, Kumar DG, Gopalakrishnan VK. Amelioration of nitrobenzene-induced nephrotoxicity by the ethanol extract of the herb *Euphorbia hirta*. *Phcog Res* 2011;3:201-7.

Source of Support: Nil, **Conflict of Interest:** None declared.