

Acaricidal effect of an isolate from *Hoslundia opposita* vahl against *Amblyomma variegatum* (Acari: Ixodidae)

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ABSTRACT

Background: *Hoslundia opposita* Vahl. (Lamiaceae), a common local shrub in Ghana, is traditionally known not only for its pharmacological benefits but also for its insecticidal properties. Its acaricidal property, however, has not been investigated. **Objective:** To test the acaricidal effects of the crude extract and fractions of *H. opposita* leaves as well as to isolate and characterize the acaricidal principles. **Materials and Methods:** The crude methanolic extract, pet. ether, ethyl acetate and aqueous fractions of the leaves of *H. opposita* were tested against the larvae of the cattle tick, *Amblyomma variegatum*, using the Larval Packet Test. A bioassay-guided isolation was carried out to identify the acaricidal principle obtained from the ethyl acetate fraction. **Results:** The active principle was characterised as ursolic acid, a triterpene previously isolated from the leaves of the same plant. The extract and fractions were less potent than the control, malathion (LC_{50} 1.14×10^{-4} mg/ml). Among the plant samples however the crude methanolic extract exhibited the highest effect against the larvae (LC_{50} 5.74×10^{-2} mg/ml), followed by the ethyl acetate fraction (LC_{50} 8.10×10^{-2} mg/ml). Ursolic acid, pet. ether and aqueous fractions however showed weak acaricidal effects with LC_{50} values of 1.13 mg/ml, 8.96×10^{-1} mg/ml and 1.44 mg/ml, respectively. **Conclusion:** Ursolic acid was not as potent as the crude methanolic extract and the ethyl acetate fraction from which it was isolated. The overall acaricidal effect of *H. opposita* may have been due to synergy with other principles having acaricidal properties.

Key words: Acaricide, Larval Packet Test, ticks, ursolic acid

INTRODUCTION

Ghana has an estimated cattle population of more than 1.5 million.^[1] It is a source of livelihood for many in the rural areas.^[2] Considered to be the greatest animal disease problem in Africa, ticks and tick-borne diseases (TDBs) are therefore a great threat and concern to cattle owners in Ghana.^[3,4]

A myriad of problems are caused by ticks, notably disease transmission, reduction of animal growth rate, milk yield, livestock productivity and market opportunities.^[5] Ticks and TBDs have substantial control costs, requiring considerable expenditures to treat, limit their spread and protect productivity.^[3,5]

The use of chemical acaricides in tick control is associated with a number of problems, including environmental pollution, chemical residues in meat, wool and milk products, toxicity to farm animals, high cost of chemicals, pesticide poisoning and the rapid development of tick resistance.^[3,6]

The search for safer and effective alternatives has soared and successes have been obtained through natural products, especially those from plant sources.^[7,8] They are economically and environmentally friendly and less-toxic to humans and animals.

Hoslundia opposita Vahl. (fam. Lamiaceae) is a common perennial shrub that survives well in both wet and dry areas of Ghana. It has been shown to have several pharmacological benefits, both traditionally and scientifically. Essential oils from the leaves are extensively used as insect repellents. Although *H. opposita* has been widely studied for many pharmacological activities, its acaricidal potential, however, is yet to be sourced.

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This study therefore seeks to investigate the acaricidal potential of *H. opposita* leaf extract against a common cattle tick in Ghana, *Amblyomma variegatum*.

MATERIALS AND METHODS

Collection and authentication of plant material

Fresh leaves of *H. opposita* were collected from a farmland at Juaben, in the Ashanti region of Ghana. It was authenticated at the Department of Pharmacognosy, Faculty of Pharmacy and Pharmaceutical Sciences, Kwame Nkrumah of Science and Technology, Kumasi, Ghana, where a voucher specimen (HO1/2010) has been deposited at the department's Herbarium.

Extraction and isolation of the active acaricidal Principle

Approximately 1.0 kg of the air-dried leaves was cold-macerated with 5.0 L of methanol for 7 days. With the aid of the rotary evaporator, the extract was concentrated under reduced pressure and further dried to give a solid mass of 90.9 g (9.09% w/w).

Subsequently, 50 g of the crude methanolic extract (CME) was partitioned between 5×100 ml aliquots of petroleum ether (40–60°) and water [1:1]. The petroleum ether fraction was collected, concentrated into a solid friable mass of 23.7 g and designated F1. Then, 5×100 ml aliquots of ethyl acetate were shaken with the aqueous extract. The ethyl acetate fraction was also collected and concentrated into a solid friable mass of 5.5 g and designated F2. The aqueous fraction left (F3) was then concentrated and freeze-dried to yield 10.0 g of amorphous mass.

Five grams of the ethyl acetate fraction (active) was repeatedly column-chromatographed on silica gel 60 F254 (70–230 mesh ASTM), using pet. ether–ethyl acetate step gradient. Guided by their thin layer chromatography (TLC) profiles, fractions collected were bulked into four main fractions labeled A to D. Fraction C (active) was repeatedly column-chromatographed on silica gel 60 F254 (70–230 mesh ASTM), employing gradient elution with pet. ether–ethyl acetate mixture (increasing polarity). Eluates collected were bulked into three main fractions, C1 to C3. Fraction C2 (active) afforded an off-white amorphous powder, X (78 mg), after washing contaminants off with 5% ethyl acetate in pet. ether.

Bioactivity studies

Biological material

Engorged adult female ticks of *A. variegatum* were obtained from the Kumasi Abattoir and bred for eggs. The 5-day

old, unfed larvae were used in this experiment.

Bioactivity studies of extracts and fractions

The crude extract and fractions were screened for acaricidal effect using the Larval Packet Test (LPT). Twenty 5-day old, unfed *A. variegatum* larvae were brought into contact with drug-impregnated filter papers (Watman No. 1) and observed for mortality after 24 h. A larva was recorded dead if it showed no sign of mobility. Concentrations used were serially diluted and ranged from 5.0 to 5.0×10^{-5} % w/v. A broad-spectrum acaricide, malathion, was used as the positive control, while methanol was used as the negative control. All experiments were run in triplicate.

Acaricidal activity of the isolate

Concentrations ranging from 2.0 to 2.5×10^{-10} % w/v were prepared by serial dilutions. These were tested against the 5-day-old, unfed larvae as described above, using methanol and malathion as negative and positive controls, respectively.

Statistical analysis

Results were expressed as LC_{50} values compared with the control. One-way ANOVA was used for comparison of the means.

RESULTS AND DISCUSSIONS

A number of ethnomedicinally important plant species abound in Ghana. Essential oils from such plants are commonly used as repellents or pesticides. Traditional livestock farmers employ various herbs to protect their animals and themselves against ticks and TBDs. Although research into local acaricidal plants has been scanty, one study by Annan *et al.*^[9] that investigated the acaricidal effects of a common shrub, *Plumbago zeylanica*, identified the naphthoquinone, plumbagin, as the main acaricidal principle. Their finding therefore suggests the possibility of many more acaricidal principles lying hidden in many local plant species used as acaricides.

Isolate X was isolated as an off-white amorphous powder with an R_f value of 0.38 in pet. ether:ethyl acetate (8:2) and 0.67 in chloroform:methanol (9:1). It recorded a melting point of 270–273°C (uncorrected). ESI-TOF-MS data of X showed a molecular ion peak $[M^+]$ at m/z 456.4, which corresponds to the molecular formula $C_{30}H_{48}O_3$.

¹H-NMR spectral data showed that X has a vinylic proton triplet at δ 5.14 and an alcoholic proton triplet at δ 3.09. It also showed an allylic proton multiplet at δ 1.87 and several saturated alkane protons with chemical shifts ranging from δ 0.63 to δ 1.52.

¹³C-NMR spectrum showed the presence of 30 carbon signals. The DEPT 135 spectrum showed seven methyl groups at δ15.25, δ15.45, δ16.73, δ16.85, δ20.98, δ23.37 and δ27.89. Nine methylene groups were shown at δ18.20, δ23.15, δ24.09, δ26.67, δ29.55, δ30.56, δ32.93, δ36.71 and δ38.57, while seven methine groups were recorded at δ38.80, δ38.98, δ47.45, δ52.72, δ55.15, δ78.70 and δ125.40. Seven quaternary carbons were also shown at δ36.81, δ39.09, δ39.35, δ41.95, δ47.70, δ138.08 and δ180.61. X was shown to possess a carboxylic acid carbon at δ180.61, two olefinic carbons at δ125.40 and δ138.08 and a saturated oxygen-bearing methine at δ78.70. The MS, ¹H-NMR and ¹³C-NMR spectral data compared closely with that of ursolic acid as reported by the literature^[10] [Table 1]. The structure of isolate X was therefore assigned as ursolic acid [Figure 1], previously isolated from the leaves of the same species.^[11]

Biological activity

The crude methanolic extract was the most potent among the test samples against *A. variegatum* larvae (LC₅₀ 5.74 × 10⁻² mg/ml). The ethyl acetate fraction also showed a relatively high potency (LC₅₀ 8.10 × 10⁻² mg/ml). The pet.

ether and aqueous fractions showed weak acaricidal effects, recording LC₅₀ values of 8.96 × 10⁻¹ mg/ml and 1.44 mg/ml, respectively. Ursolic acid also exhibited a weak acaricidal effect against the larvae of *A. variegatum*. At 2.0% w/v, it showed 58.5% mortality, with an LC₅₀ value of 1.13 mg/ml. This was significantly lower (*P* < 0.001) than the effect exhibited by the control, malathion (LC₅₀ 1.14 × 10⁻⁴ mg/ml) [Table 2].

Malathion is a broad-spectrum acaricide commonly used in Ghana. It is one of several others used by livestock farmers. However, due to the high cost associated with its use,^[2] most resource-poor farmers have resorted to cheaper alternative means of arriving at similar outcomes. One such alternative is their traditional knowledge and practices of herbal acaricides. Oils from various parts of plants have been used to repel or kill pests, including seeds, fruit peels and flowers.^[12,13]

Ursolic acid is a triterpene, a known component of many essential oils. Literatures supporting the ethnomedicinal use of essential oils in pest control continue to increase. These effects have been attributed to the terpene components of these plants.^[13,14]

Table 1: ¹H-NMR and ¹³C-NMR spectral data for ursolic acid in CDCl₃/CD₃OD

Position	Chemical shift (ppm)		Reference ^[10]	
	¹ H-NMR	¹³ C-NMR	¹ H-NMR	¹³ C-NMR
C-1		38.57		38.90
C-2	3.09	26.67	3.11	27.10
C-3	-	78.70	-	79.00
C-4	0.63	39.09	-	39.00
C-5	1.21	55.15	-	55.60
C-6	-	18.20	-	18.60
C-7	-	32.93	5.16	33.40
C-8	5.14	39.35	-	39.80
C-9	-	47.45	-	47.90
C-10	-	36.81	-	37.20
C-11	-	23.15	2.11	23.60
C-12	2.10	125.40		125.80
C-13	1.52	138.08	0.90	138.50
C-14		41.95	0.74	42.40
C-15	0.72	29.55	0.70	29.90
C-16	0.68	24.09	0.84	24.50
C-17	1.00	47.70	1.01	48.10
C-18	-	52.72	-	53.20
C-19	0.87	38.98	0.86	39.40
C-20	0.77	38.80	0.78	39.20
C-21		30.56		30.90
C-22		36.71		37.10
C-23		27.88		28.30
C-24		15.25		15.60
C-25		15.45		15.80
C-26		16.73		17.20
C-27		23.37		23.70
C-28		180.61		180.80
C-29		16.85		17.10
C-30		20.98		21.30

ppm: parts per million

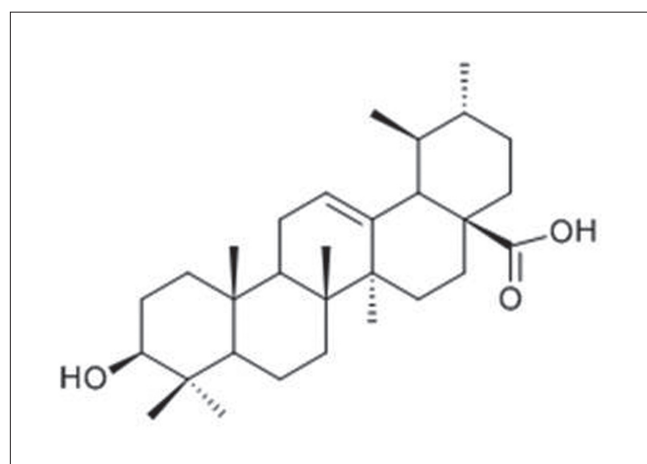


Figure 1: Structure of Ursolic acid

Table 2: Acaricidal effects of crude extract and fractions of *H. opposita* leaves against larvae of *A. variegatum* compared with control

Material	LC ₅₀ (mg/ml)	Range of % mortality (%)
Crude methanolic extract	5.74 × 10 ⁻²	8.5–100.0
Pet. ether fraction	8.96 × 10 ⁻¹	6.5–45.0
Ethyl acetate fraction	8.10 × 10 ⁻²	13.5–95.0
Aqueous fraction	1.44	0.0–13.5
Ursolic acid	1.13	8.5–58.5
Malathion	1.14 × 10 ⁻⁴	38.5–100.0

LC₅₀: concentration required to kill 50% of the test organisms, mg/ml: mass in milligrams per 1 ml, %: percent

Acaricidal effects may have been higher in the crude methanolic extract and the ethyl acetate fraction due to the fact that there was a combined effect by other components of the extract and fraction. Previous phytochemical screening of *H. opposita* showed the presence of other medicinally important terpenes like menthol, thymol, β -caryophyllene, oleanolic acid and germacrene-D. A study has even reported on the presence of benzyl benzoate, a known pesticide^[15] identified from samples of essential oils obtained from the plant in West Africa. The combined effects of components such as these may have contributed to the high effects observed. Some structure–activity studies on terpenes suggest that molecules possessing free alcoholic, phenolic or potential olefinic modifications showed the most potent acaricidal activity.^[16,17]

CONCLUSION

The results of this study suggest that the crude methanolic extract and ethyl acetate fraction of the leaves of *H. opposita* have acaricidal activity against the larvae of *A. variegatum*. Ursolic acid has been found to be a component of the ethyl acetate fraction, which contributes to the acaricidal activity of the plant. The finding therefore corroborates the traditional use of the plant as a repellent; however, its great potential would be limited by using it only as a repellent. The alcoholic extract of the leaves can serve the livestock industry better as a safer and cheaper alternative to chemical acaricides.

REFERENCES

1. Kaljouw M. Resistance to acaricides of *Boophilus* ticks from cattle in Ghana. PhD Thesis. The Netherlands: Utrecht University; 2009.
2. Minjauw B, McLeod A. Tick-borne diseases and poverty. The impact of ticks and tick-borne diseases on the livelihood of small-scale and marginal livestock owners in India and eastern and southern Africa. Research report, DFID Animal Health Programme, Centre for Tropical Veterinary Medicine, University of Edinburgh, UK; 2003.
3. Kaaya GP. Prospects for Innovative Tick Control Methods in Africa. *Insect Sci Appl* 2003;23:59-67.
4. Bell-Sakyi L, Koney EB, Dogbey O, Walker AR. Incidence and prevalence of tick-borne haemoparasites in domestic ruminants in Ghana. *Vet Parasitol* 2004;124:25-42.
5. McDermott J, Richard D, Randolph T. Incidence of animal diseases and their current and future impacts on crop-livestock systems in West Africa. In: Williams TO, Tarawali S, Hiernaux P, Fernandez-Rivera S, editors. Sustainable crop-livestock production for improved livelihoods and natural resource management in West Africa. Proceedings of an international conference; 2001 Nov. 19-22. ILRI, Nairobi (Kenya) and CTA, Wageningen, The Netherlands; 2004.
6. Miller GT. *Sustaining the Earth*. 6th ed. California: Thompson Learning Inc, Pacific Grove; 2004.
7. De Boer H, Vongsombath C, Pålsson K, Björk L, Jaenson TG. Botanical repellents and pesticides traditionally used against hematophagous invertebrates in Lao people's democratic republic: A comparative study of plants used in 66 villages. *J Med Entomol* 2010;47:400-14.
8. Rasikari HL, Leach DN, Waterman PG, Spooner-Hart RN, Basta AH, Banbury LK, *et al.* Acaricidal and Cytotoxic Activities of Extracts from Selected Genera of Australian Lamiaceae. *J Econ Entomol* 2005;98:1259-66.
9. Annan K, Dickson R, Mensah A, Fleischer TC. Acaricidal Effect of *Plumbago zeylanica* L. against *Amblyomma variegatum* Pharmacogn J 2009;1:190-4.
10. Nthambeleni R. Isolation and characterisation of cytotoxic compounds from *Anthospermum hispidulum* and *Eriocephalus tenuifolius*. MSc. Thesis. Pietermaritzburg, SA: University of KwaZulu-Natal; 2008.
11. Ngadjui BT, Tsopmo A, Ayafor JF, Connolly JD, Tamboue H. Hosloppin, a New Pyrone-substituted flavonoid from *Hoslundia opposita*. *J Nat Prod* 1995;58:109-11.
12. Dolan MC, Dietrich G, Panella NA, Montenieri JA, Karchesy JJ. Biocidal Activity of three wood essential oils against *Ixodes scapularis* (Acari: Ixodidae), *Xenopsylla cheopis* (Siphonaptera: Pulicidae), and *Aedes aegypti* (Diptera: Culicidae). *J Econ Entomol* 2007;100:622-5.
13. Han J, Choi BR, Lee SG, Kim SI, Ahn YJ. Toxicity of plant essential oils to acaricide-susceptible and -resistant *Tetranychus urticae* (Acari: Tetranychidae) and *Neoseiulus californicus* (Acari: Phytoseiidae). *J Econ Entomol* 2010;103:1293-8.
14. Cseke LJ, Kirakosyan A, Kaufman SL, Duke JA, Brielman HL. *Natural Products from Plants*. 2nd ed. Boca Raton: Taylor and Francis Group; 2006.
15. Tonzibo ZF, Ahibo CA, Chalachat JC, N'guessan YT. Chemical composition of essential oils of *Hoslundia opposita* Vahl. from Ivory Coast. *Flavour Fragr J* 2006;21:789-91.
16. Perrucci S, Macchioni G, Cioni PL, Flamini G, Morelli I. Structure/activity relationship of some natural monoterpenes as acaricides against *Psoroptes cuniculi*. *J Nat Prod* 1995;58:1261-4.
17. Cantrell CL, Klun JA, Pridgeon J, Becnel J, Duke S. Structure/Activity relationship studies on the arthropod repellent callicarpenal. USDA-ARS; 2009. Available from: <http://www.ars.usda.gov>. [Last accessed on 2010 Oct 13].

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